

## METHODOLOGY FOR DETERMINATION OF PLASMA CORTISOL IN FISH USING COMPETITIVE ENZYME-LINKED IMMUNOSORBENT ASSAY (ELISA)

### METODOLOGÍA PARA LA DETERMINACIÓN DE CORTISOL PLASMÁTICO EN PECES USANDO LA PRUEBA DE INMUNOENSAYO ENZIMÁTICO (ELISA)

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#### ABSTRACT

**Objective.** To determine plasma cortisol procedure in fish using competitive enzyme-linked immunosorbent assay (ELISA). **Materials and methods.** Two plasma samples of juveniles rainbow trout *Oncorhynchus mykiss* were analyzed by using ELISA human kit for cortisol assay. For standard curve calibration seven standard solutions of cortisol in human plasma (0, 20, 50, 100, 200, 400 and 800 ng.ml<sup>-1</sup>) were used. For the recovery test 50, 100 and 200 ng.ml<sup>-1</sup> standard solutions of cortisol were used; for the linearity test four dilutions of the fish plasma samples (1/2, 1/4, 1/8 and 1/16) were prepared. To each well fish plasma samples and standard solutions were added and its content were conjugated to peroxidase and subsequently enzyme substrate was added. The enzymatic reaction was stopped by addition of phosphoric acid 0.5 M and the absorbance was read at 450 nm. The accuracy of the pipetting procedure was assessed previously. The recovery and linearity percentages, the standard curve and parallelism were determined. **Results.** The standard curve showed a high correlation coefficient ( $r^2 = 0.998$ ). The cortisol concentration of two samples fluctuated between 64 and 72 ng.ml<sup>-1</sup>. Only the 200 ng.ml<sup>-1</sup> standard solution showed a recovery percentage superior to 80%; in contrast, in 50 and 100 ng.ml<sup>-1</sup> the recovery percentage fluctuated between 52 and 71%. In the dilution of 1/2 to 1/8 were observed a good linearity (86 to 168%); the samples showed parallelism with the standard curve. **Conclusions.** The use of human plasma cortisol for ELISA procedure is an accuracy and efficiency test for fish cortisol plasma determination.

**Key words:** Cortisol, ELISA, fish, *Oncorhynchus mykiss*, rainbow trout, plasma.

## RESUMEN

**Objetivo.** Describir el procedimiento para determinar cortisol plasmático en peces, utilizando la prueba de inmunoensayo enzimático (ELISA). **Materiales y métodos.** Dos muestras de plasma de trucha arco iris *Oncorhynchus mykiss* fueron analizadas empleando un kit de ELISA desarrollado para humanos. Siete soluciones estándar conteniendo 0, 20, 50, 100, 200, 400 y 800 ng.ml<sup>-1</sup> de cortisol fueron usadas para construir una curva de calibración. Para la prueba de recuperación se emplearon las soluciones estándar de 50, 100 y 200 ng.ml<sup>-1</sup>; finalmente, para la prueba de linealidad se prepararon cuatro diluciones de las muestras de plasma, así: 1/2, 1/4, 1/8 y 1/16. A cada pozo de la placa se adicionaron tanto las muestras de plasma como las soluciones estándar, las cuales fueron conjugadas con peroxidasa y posteriormente se adicionó el substrato de la enzima. Esta reacción enzimática se detuvo por medio de la adición de ácido fosfórico (0.5 M) y posteriormente, la absorbancia fue medida a 450 nm. La precisión del procedimiento de pipetaje fue evaluado previo a la prueba. El porcentaje de recuperación y de linealidad, así como la curva de calibración y de paralelismo fueron determinadas. **Resultados.** La curva estándar mostró un alto coeficiente de correlación ( $r^2 = 0.998$ ). La concentración de cortisol en las dos muestras de plasma fluctuó entre 64 y 72 ng.ml<sup>-1</sup>. Sólo la solución estándar de 200 ng.ml<sup>-1</sup> mostró un porcentaje de recuperación superior al 80%; en contraste, en las soluciones estándar de 50 y 100 ng.ml<sup>-1</sup> el porcentaje de recuperación fluctuó entre 52 y 71%. En las diluciones de 1/2 y 1/8 se observó un buen porcentaje de linealidad (86 a 168%). Finalmente, las muestras mostraron cierto grado de paralelismo con la curva estándar. **Conclusiones.** El uso de la prueba de ELISA para determinar cortisol plasmático en humanos, es confiable y eficiente para la cuantificación de cortisol plasmático en peces.

**Palabras clave:** Cortisol, ELISA, *Oncorhynchus mykiss*, peces, trucha arco iris, plasma.

## INTRODUCTION

There are three different stress responses in teleosts including primary, secondary and tertiary response, each of one with several physiological mechanisms (1). In the primary responses, cortisol is the principal corticosteroid identified in teleosts (2), being the most sensitive steroid hormone marker (1) produced in the interrenal cells by the ACTH stimulation and released in response to a variety of stressors (3, 4). The main responses influenced by stress include effects on ionic and osmotic regulation with changes in the plasma ionic concentration, branchial ionic fluxes, plasma osmotic pressure, volume and composition of the urine and

salt absorption (5); protein and carbohydrate metabolism, stimulating gluconeogenesis from amino acids and glycogen in the liver and thus increasing blood glucose (1, 6); blood cell movement from haematopoietic and lymphoid tissues (7); immune system with disturbance in tissue repair, anti-inflammatory reactions, phagocytosis and lymphoid system (7,8) and effects on growth (7).

Cortisol has been measured using enzyme-linked immunosorbent assay (ELISA), one of the most suitable immunoassay techniques owing to high sensitivity and specificity (1). In addition, this technique does not require expensive instrumentation or advanced

technical expertise (9,10); perhaps, the most significant advantage on others immunoassays such as radioimmunoassay (RIA) is that ELISA does not need radioisotopes (10). In fish the use of direct ELISA is the most common (11, 12); however, Tintos et al (1) validated the use of indirect ELISA assay to determine plasma cortisol in fish. Therefore, the aim of the present study was describe the methodology of the competitive ELISA to measure plasma cortisol in juvenile rainbow trout *Oncorhynchus mykiss* (Walbaum).

## MATERIALS AND METHODS

**Pipetting calibration.** Pipetting volume of 20, 50, 100 and 200  $\mu\text{l}$  were weighed in analytical balance in order to guaranty the accuracy of the pipetting procedure during the ELISA assay. Each volume taken was weighed in a plastic container with a previous zeroing of the balance. Table 1 shows the weight of each pipetting volume.

**ELISA assay for cortisol.** Two juveniles of rainbow trout *Oncorhynchus mykiss* with *c.a.* 15 - 20 cm of fork length and 90 - 150 g of body weight were anaesthetized by immersion in MS-222 solution (0.1%, Sigma Co., St Louis, Missouri). Blood samples were taken from the caudal peduncle using heparinized syringes to obtain plasma after centrifugation at 10,000  $\times g$  for 5 min, maintained on ice until determination of cortisol concentrations.

Ninety-six well plates for cortisol ELISA (DRG Diagnostics, Frauenbergstrasse, Germany) were used. For this assay only 38 wells were used and the fish plasma samples were analysed in duplicate.

**Recovery test.** Three Eppendorf tubes of 1.5 mL were prepared with standard

solution of 50, 100 or 200  $\text{ng}\cdot\text{ml}^{-1}$  and fish plasma samples. Standard solutions and fish plasma samples were mixed 1:1 (v:v) and to each tube previously labelled were added 30  $\mu\text{l}$  of fish plasma sample and 30  $\mu\text{l}$  of the respective standard solution. The tubes were maintained at room temperature (*c.a.* 20°C).

**Linearity test.** Four Eppendorf tubes (1/2, 1/4, 1/8, 1/16) indicating the dilution of the fish plasma sample were labelled; in this case standard solutions and fish plasma samples were mixed 1:1 (v:v). For the first dilution (1/2) 30  $\mu\text{l}$  of fish plasma sample and 30  $\mu\text{l}$  of standard 0  $\text{ng}\cdot\text{ml}^{-1}$  were mixed. Consequently, 30  $\mu\text{l}$  of the 1/2 dilution and 30  $\mu\text{l}$  of 0  $\text{ng}\cdot\text{ml}^{-1}$  standard solution was taken and mix in the second tube to prepare the dilution of 1/4. This procedure was repeated for the two remainder dilution. The samples were maintained at room temperature.

**Assay protocol.** For the assay, 20  $\mu\text{l}$  of each of cortisol human plasma standard solution (0, 20, 50, 100, 200, 400 and 800  $\text{ng}\cdot\text{ml}^{-1}$ ) and fish plasma sample were added in duplicate to the plate. In the same way, the samples for the recovery and linearity test were dispensed in others wells. Standard solutions and fish plasma samples for recovery test were assayed in duplicate.

Subsequently, 200  $\mu\text{l}$  of enzyme conjugated to horseradish peroxidase (DRG Diagnostics, Frauenbergstrasse, Germany) was added into each well. Finally, the wells were gently mixed on a plate mixer at a 200  $\text{beats}\cdot\text{min}^{-1}$  for 10 min and incubated for 1 h at room temperature.

The well contents were briskly eliminated to avoid any residual content. The solution of each well was removed by

washing the plate three times with 400  $\mu$ l of PBS and shaking out the content onto absorbent paper with the aim of removing residual drops that could affect the accuracy and precision of the assay. Subsequently, 100  $\mu$ l of TMB (tetramethylbenzidine) enzyme substrate (DRG Diagnostics, Frauenbergstrasse, Germany) was added to each well and incubated for 15 min at room temperature. The enzymatic reaction was visualized by the colour change and was stopped by addition of 100  $\mu$ l of 0.5 M phosphoric acid ( $H_2PO_3$ ). The intensity of colour is inversely proportional to the concentration of cortisol in the samples. Absorbance was read in a spectrophotometer at 450 nm on a microtiterplate reader within 10 min after addition of stop solution.

**Curves.** A standard curve was plotted with the aim of calculating the concentration of cortisol of each plasma sample. The curve was plotted with log concentration of each standard solution against logit optical density. For this, the mean of the absorbance values of 20, 50, 100, 200, 400 and 800  $ng \cdot ml^{-1}$  standard solutions was used. The optical density (OD) was calculated as absorbance of the each standard solution

over the absorbance of the zero standard solution. Once OD was determined, the logit OD was calculated using the follow equation: Logit OD:  $\log (OD/(100-OD))$ .

Finally, with the aim to verify the parallelism between standard curve and fish diluted plasma samples, the logarithm relative (log relative) of the fish diluted plasma samples against logit OD of these diluted samples were plotted in the same graph with the standard curve.

All values are expressed as mean  $\pm$  SEM. For all statistical analysis, GraphPad InStat Software version 3.06 (1992-2003) and GraphPad Prism Software version 4.03 (1992-2005) were used.

## RESULTS

The accuracy of the pipetting procedure was observed in the four pipetting volumes evaluated (Table 1). Table 2 summarises the results of optical density (OD) and logit OD of the standard solutions. Most of the values of absorbance for standard solutions showed a coefficient of variance (CV) less than 10%; however, the 400  $ng \cdot ml^{-1}$  standard solution showed a high CV (18%).

**Table 1.** Pipetting accuracy of different volumes using an analytical balance to ensure the technique.

Replicate	Weight (mg)			
	20 $\mu$ l	50 $\mu$ l	100 $\mu$ l	200 $\mu$ l
1	21.3	49.7	98.8	199.3
2	20.3	50.1	99.4	203.6
3	19.9	49.2	98.8	200.3
4	20.3	49.6	99.3	200.0
5	20.1	49.3	99.8	201.1
6	20.0	49.6	99.1	200.1
Mean	20.3 $\pm$ 0.2	49.6 $\pm$ 0.1	99.2 $\pm$ 0.2	200.7 $\pm$ 0.6

The values are expressed per replicate ( $n = 6$ ). The final row shows the values as a mean  $\pm$  SEM.

**Table 2.** Procedure for determination of optical density (OD) and logit OD of standard solutions values of plasma cortisol assay.

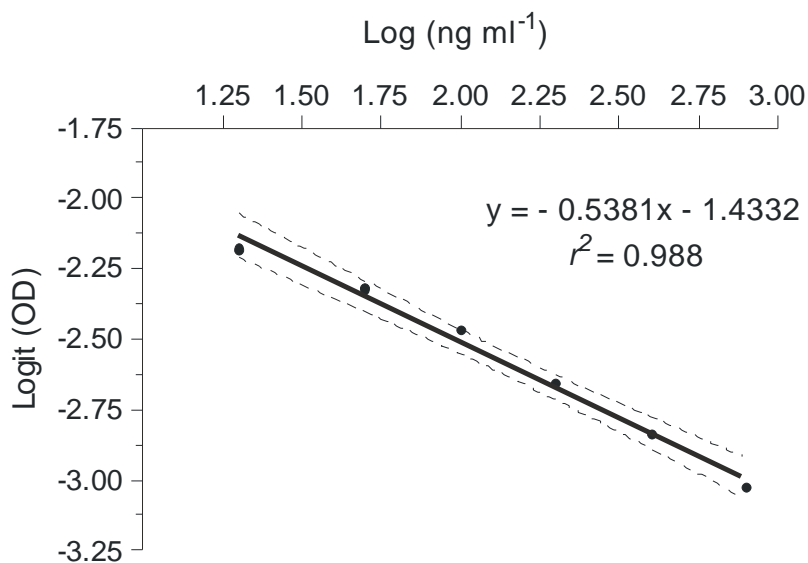
Standard solutions (ng ml <sup>-1</sup> )	Absorbance (nm)	OD <sup>1</sup>	Logit (OD) <sup>2</sup>
0	1.135 ± 0.045 (5.61)	—	—
20	0.744 ± 0.023 (4.46)	0.656	-2.180
50	0.538 ± 0.005 (1.44)	0.474	-2.3217
100	0.384 ± 0.009 (3.31)	0.338	-2.469
200	0.249 ± 0.017 (9.92)	0.220	-2.657
400	0.166 ± 0.021 (17.89)	0.146	-2.834
800	0.106 ± 0.005 (7.3)	0.094	-3.027

The values are expressed as mean ± SEM. The coefficient of variance is show in parenthesis.

The standard curve showed a high correlation coefficient between logit OD and log of standard solution concentration (Figure 1). According to the equation obtained with the standard

curve ( $Y = -0.5381x - 1.4332$ ), the equation for cortisol determination was calculated as follows:

$$X = (\text{logit OD} + 1.4332) / -0.5381$$

**Figure 1.** Standard curve for cortisol determination using logit OD against log of standard cortisol solution concentration ( $r^2 = 0.988$ ).

Where X corresponds to the cortisol concentration expressed as a logarithm; then, the antilogarithm of X value was calculated to obtain the final concentration of cortisol. The procedure for calculation of the cortisol concentration is summarised as follows: the absorbance

obtained for each sample was used to obtain the OD; both OD and logit OD were calculated as described above. In the two plasma fish samples a low CV was observed. The cortisol concentration of two fish samples fluctuated between 64 and 72 ng.ml<sup>-1</sup> (Table 3).

**Table 3.** Procedure for determination of optical density (OD) and logit OD of plasma samples of rainbow trout (*Oncorhynchus mykiss*).

Sample	Absorbance (nm)	OD <sup>1</sup>	Logit (OD) <sup>2</sup>	Logarithm of cortisol (X) <sup>3</sup>	[Cortisol] (ng ml <sup>-1</sup> ) <sup>4</sup>
1	0.418 ± 0.029 (9.97)	0.368	-2.432	1.856	71.71
2	0.446 ± 0.032 (10.15)	0.393	-2.404	1.804	63.68

<sup>1</sup>OD = absorbance of the sample/absorbance of zero standard (1.135) - <sup>2</sup>Logit OD = log (OD / (100/OD)) - <sup>3</sup>X = (logit (OD) + 1.4332) / - 0.5381 - <sup>4</sup>[cortisol] = 10<sup>logarithm of cortisol</sup>. The values are expressed as mean ± SEM. The coefficient of variance is show in parenthesis.

The final concentration of cortisol and recovery percentage is shows in the Table 4. Only the 200 ng.ml<sup>-1</sup> standard solution showed a recovery percentage superior to 80%; in the remainder concentrations (50 and 100 ng.ml<sup>-1</sup>) the recovery percentage fluctuated between 52 and 71%. On the other hand, in the

dilution of fish plasma sample 1/2, 1/4 and 1/8 were observed a good percentage of linearity (86 to 168%); however, the dilution of 1/16 showed the greatest value (superior to 280%, Table 5). Finally, the fish plasma samples showed a good parallelism with the standard curve (Figure 2).

**Table 4.** Cortisol concentration and recovery test of plasma samples of rainbow trout (*Oncorhynchus mykiss*).

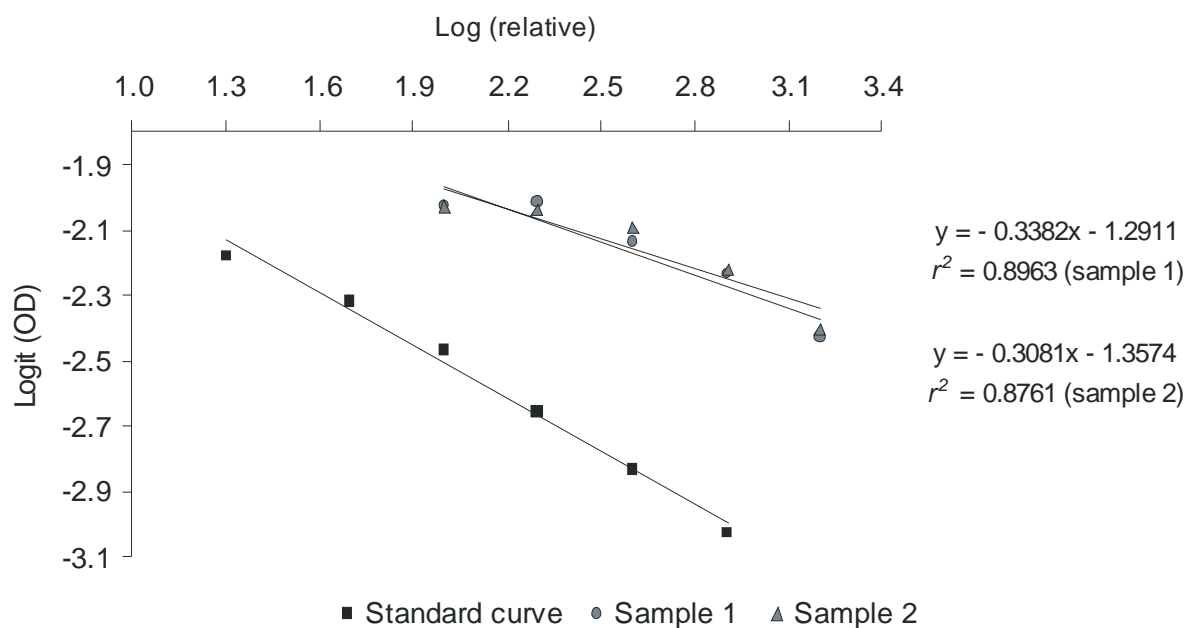
Sample	Added concentration (1:1, v:v) (ng ml <sup>-1</sup> )	Measured [cortisol] (ng ml <sup>-1</sup> )	Expected <sup>1</sup> [cortisol] (ng ml <sup>-1</sup> )	Recovery (%)
1	X	71.71	71.71	100
	50	50.32	96.71	52
	100	86.92	121.71	71.4
	200	154.76	171.71	90.1
2	X	63.68	63.68	100
	50	46.07	88.68	51.9
	100	74.34	113.68	65.4
	200	131.75	163.68	80.5

<sup>1</sup> Expected [cortisol] = (measured cortisol of X + added concentration) / 2

**Table 5.** Linearity test of plasma cortisol of rainbow trout (*Oncorhynchus mykiss*).

Sample	Dilution	Measured [cortisol] (ng ml <sup>-1</sup> )	Expected <sup>1</sup> [cortisol] (ng ml <sup>-1</sup> )	Recovery (%)
1	0	71.713	71.713	100
	1/2	30.895	35.865	86.2
	1/4	20.457	17.928	114.1
	1/8	12.353	8.964	137.8
	1/16	12.843	4.482	286.5
2	0	63.685	63.685	100
	1/2	29.292	31.842	92.0
	1/4	17.253	15.921	108.4
	1/8	13.363	7.961	167.9
	1/16	12.958	3.980	325.6

<sup>1</sup>Expected [cortisol] = expected cortisol of previous dilution / 2

**Figure 2.** Parallelism curve.

The large line corresponds to standard curve. The short lines correspond to linearity test of diluted plasma samples of juveniles rainbow trout (*Oncorhynchus mykiss*).

## DISCUSSION

The basal cortisol plasma levels in rainbow trout fluctuate between 2 to 25 ng.ml<sup>-1</sup> (1,10). Due to cortisol being the most readily stress hormone to assess the activity of hypothalamic-pituitary-

interrenal axis - HPI - (4), the increase of plasma levels has been frequently associated to short-term stressors (13).

Several factors affect the plasma cortisol levels in fish including size and age of fish, time of day, rearing temperature (10, 14,

15), reproductive stage (16), acclimation conditions, handling (11, 13), tank confinement (10, 13), use of anaesthetics (17), changes of osmotic condition (12), air exposition (11, 13) and fish strains (16). Variation of response to stressful condition among species has been reported, being salmonids one of the species that respond almost immediately to handling and crowing stress (11, 16). In the present study, the higher values of plasma cortisol (three-fold) observed could be associated with these later conditions, especially with the handling stress.

The coefficient of variance, smaller than 10%, could reflected a good level of reproducibility; however, the low number of individual sampled did not offered a high level of accuracy, being necessary a major number of repetitions.

The low percentage of recovery can not be attributed to inaccuracy of pipetting itself owing to the fact that the volume was verified correctly on the analytical balance; however, it is probably that during the pipetting method some errors have occurred. Although the procedure was done at room temperature, is probable that the bound enzyme could be affected by temperature variations and edge effects. On the other hand,

due to washing procedure affecting greatly the sensitivity and precision of the assay, is probable that the contents had not been removed correctly and this affected the recovery percentage.

The good percentage of linearity reflects a high sensitivity and affinity of this ELISA human kit to determine plasma cortisol in fish; this result was confirmed by the good parallelism observed between standard curve and fish samples dilution curve proving the validity of this assay.

In conclusion, the use of human plasma cortisol for ELISA procedure is useful for fish cortisol plasma determination and could be used as a tool to assess the stress effects in fish; however, the methodology requires high accuracy and rigour to avoid errors in the results and interpretation.

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